

# The Effects of Electrical Pulses on Structure of Cells of Plant Tissue.

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**Abstract** — the developed equivalent electrical circuit of juice-pulp mixtures and formulas for determining its electrical characteristics show that the defining element in the process of electroporation is intercellular fluid and the juice released during the crushing of the vacuoles of damaged cells.

**Index Terms** — electroporation, electric pulses, resistance, biological raw materials, cell structure

## I. INTRODUCTION

In the technology of food production associated with the extraction of juice from the plant material, an important process is the destruction of semipermeable membranes of tissue cells. On the degree of destruction semipermeable membranes of cells plant material depends juice yield and the cost of its recovery. It is also important that, after processing the cellulose cell walls did not go into the juice, as this complicates its extraction and clean up. This can be achieved by pre-treatment of plant material before extracting juice various physical methods. There are many ways to increase the permeability of the cell membranes of plant materials: mechanical, thermal, biological, and chemical processing, electrical, magnetic, acoustic, radioactive, radiation. The most common are: mechanical and thermal methods of processing of plant material. Enzymatic (biological) is rarely used because of the long duration of the enzymatic treatment. Electrophysical methods of processing are in the stage of optimization and experimental-production testing [1-4]. Promising is use for electric processing of food products short electrical pulses, whose effects on the material has a number of advantages over other electrical methods. The concentration of power and subsequent short-pulse influence on the processed material leads to a qualitatively new effects, which can be the basis for the development of new technological methods [5-8]. Also perspective is the use of combined methods of influence on plant tissue. In this case, in the first stage usually uses mechanical grinding crushing, cut into shavings, etc.

In existing technologies processing of plant raw materials are allowed considerable content of valuable components in the waste production, and existing technology recycling energy-intensive and lengthy. Therefore the maximum possible extraction of valuable components from plant material at the stage of extracting the juice from the pulp is an important task.

## II. EXPERIMENTAL

To solve this problem were investigated the effect of electrical parameters: the energy density of plasmolysis,

temperature, electric field on the electrical resistance of the juice-pulp mixture, the yield of juice, spontaneity and the overall yield of juice.

As the impacting factor used bipolar pulses with steep leading and sloping trailing front the Fig.1. As a criterion for evaluation of plasmolysis taken specific resistance of plant tissue.

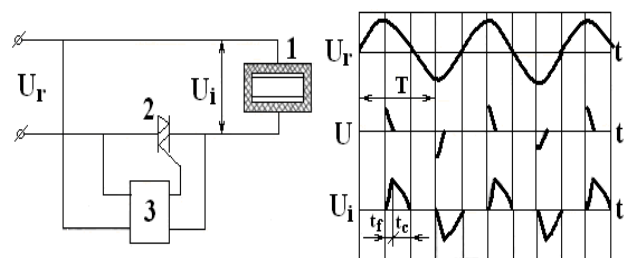


Fig.1. Circuit scheme of installation for the electroporation raw and timing diagram of voltage mains supply voltage alternating current  $U_c$ , control impulses  $U_y$  and processing pulses  $U_e$ : 1 - electrode cell, 2 - triac, 3 - the control unit.

Investigated the effects of electrical pulses on cell structure of plant tissue (Fig. 2). Microscopic and submicroscopic studies have established that under the action of electric pulses semipermeable membrane cell plasmalemma and tonoplast lose its semi-permeability and become permeable to the juice. Figure 3 shows the Microphotogram (a) and area of cross-sectional of the protoplast and the cell wall (b) of apple control sample, and in Figure 4 shows the microphotogram (a) and cross-sectional area of the protoplast and the cell wall (b) a sample of apple that been subjected by electric plasmolysis. In contrast to the control samples (Fig. 3), in the samples of processed by electrical pulses (Fig. 4) shows the significant changes. Figure 4 (b) shows a violation of the integrity of the cell protoplasm and visible breaks of the cell membrane, plasma membrane and tonoplast.

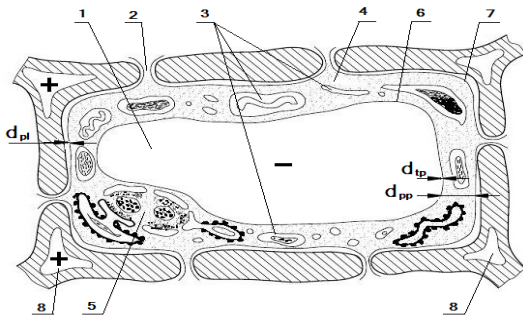


Fig.2. Schematic representation of the succulent plant cells: 1-vacuole 2 - plasmodesmata 3 - Golgi apparatus, and 4 – cytoplasm

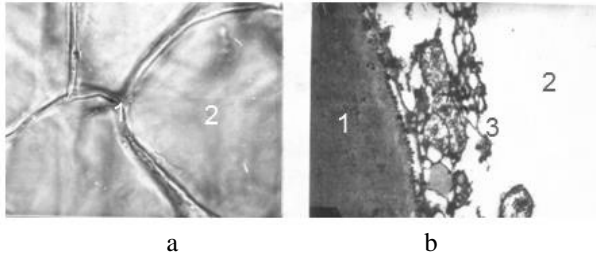


Fig.3 microphotogram (a) and cross-sectional area of the protoplast and the cell wall (b) control sample of apples: 1 - cell membrane, 2 - vacuole, 3 - tonoplast 4 - plasmalemma.

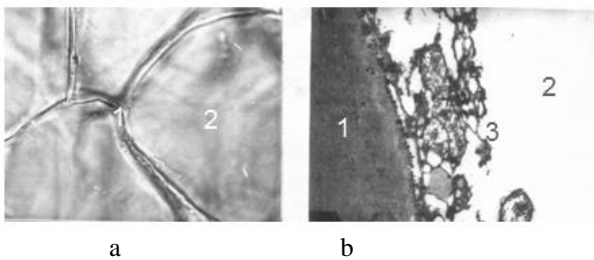


Fig.4 microphotogram (a) and cross-sectional area of the protoplast and the cell wall (b) a sample of apple subjected by electric plasmolysis: 1 - cell membrane, 2 - vacuole, 3 - fragments of cytoplasm (the tonoplast and plasmalemma).

Were developed electrical equivalent circuit, Figure 5: a) - cells; b) - tissue c) - ground tissue; d) - chopped electroplasmolysed tissue.

Living cell succulent plant materials Fig. 1 contains: vacuole -1 core - 5; cytoplasm - 4 with tonoplast - 6 and plasmalemma - 7 and intercellular spaces - 8. The crushed plant material is a piece of living plant tissue dampened with juice released from destroyed by grinding cell vacuoles

In the equivalent circuit, Figure 5:

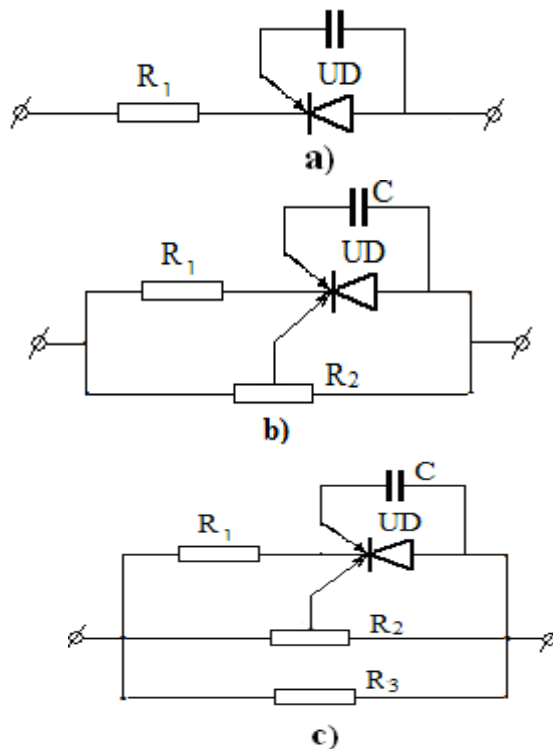
a) represented cells include: R<sub>1</sub> - resistance of vacuoles juice, C - electric capacitance the cell membrane and UD - controlled diode of semipermeable membrane cells. From the equivalent circuit of the cell can be seen that the cell under normal conditions does not permeable, and its overall resistance  $z_a = \infty$ ;

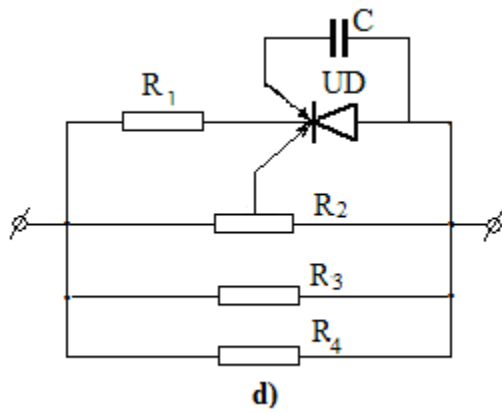
b) shows tissue plant material comprising: R<sub>2</sub> - resistance intercellular fluid and  $z_a$  - the total resistance of the cell;

from the equivalent circuit can be seen that on the total resistance of the tissue  $z_b$  in normal conditions affects only resistance of the intercellular fluid. Considering that the measured resistivity tissue plant raw material under normal conditions is approximately 16 ohms. cm, and the resistivity of the intercellular fluid around 100 Om.sm, the coefficient of concentration of interstitial fluid in the plant tissue  $k_b$  will be approximately approximately 0.006.

c) - presented ground tissue plant raw material which includes: R<sub>3</sub> - resistance juice released from the cells of tissue in the during grinding and  $z_b$  - total resistance of tissue; from equivalent circuit can be seen that on the total resistance of the ground tissue  $z_s$  affects resistance juice released from the cells tissue during grinding R<sub>3</sub> and the total resistance of tissue  $z_b$ . Considering that the measured resistivity ground tissue plant raw material is about 6 kOm.sm., and the resistivity of the cell vacuole juice about 1.2 kOm. cm, the concentration ratio of vacuoles juice in ground plant tissue  $k_c$  will be approximately 0.2.

d) - presented ground electroplasmolysed tissue plant raw material which includes: R<sub>4</sub> - resistance juice that has lost its connection with the cell as a result of electroplasmolysis ground plant material and  $z_s$  - total resistance of ground tissue; from the equivalent circuit of electroplasmolysed ground tissue can be seen that on the total resistance of the ground electroplasmolysed tissue  $z_d$  affects the resistance of juice that lost its connection with the cell as a result of electroplasmolysis of ground plant raw material and the total resistance R<sub>4</sub> ground tissue  $z_s$ . Considering that the measured resistivity of plant raw material ground electroplasmolysed tissue is about 2.5 kOm. cm, and the resistivity of the vacuoles cell juice about 1.2 kOm.sm, the concentration ratio of vacuoles juice in electroplasmolysed ground plant tissue  $k_d$  will be approximately 0.85.





$$z_a = \sqrt{R_1^2 + \left(\frac{1}{\omega C}\right)^2}; \quad z_b = \frac{\sqrt{R_1^2 + \left(\frac{1}{\omega C}\right)^2} \times R_2}{\sqrt{R_1^2 + \left(\frac{1}{\omega C}\right)^2} + R_2};$$

$$z_c = \frac{\sqrt{R_1^2 + \left(\frac{1}{\omega C}\right)^2} \times R_2 \times R_3}{\sqrt{R_1^2 + \left(\frac{1}{\omega C}\right)^2} + R_2 + R_3}; \quad z_d = \frac{\sqrt{R_1^2 + \left(\frac{1}{\omega C}\right)^2} \times R_2 \times R_3 \times R_4}{\sqrt{R_1^2 + \left(\frac{1}{\omega C}\right)^2} + R_2 + R_3 + R_4};$$

$$\rho_a \approx \infty; \quad \rho_b = \frac{\rho_e}{s_e}; \quad \rho_d = \frac{\rho_b \rho_3 \rho_4}{\rho_b + \rho_3 + \rho_4};$$

$$k_b = \frac{\rho_2}{\rho_5}; \quad k_n = \frac{\rho_1}{\rho_3}; \quad k_d = \frac{\rho_1}{\rho_4};$$

Fig.5. Equivalent electrical circuit of replacement of succulent plant raw material: a1) - cells; b1) - tissues c1) - ground tissue; d) - chopped electric electroplasmolysed tissue and formulas for determining the impedance: z1) - cells; z2) - tissue; z3) - ground tissue; z4) - chopped electric electroplasmolysed tissue and formulas for determining the resistivity: ρa) - cells; ρb) - tissue; ρc) - ground tissue; ρd) - chopped electric electroplasmolysed tissue. Where resistance: R1 - cell sap vacuole; R2 - interstitial fluid; R3 - juice after grinding of raw materials; R3 - juice after electroplasmolysis of raw materials, and measurement of resistivity: ρ1 – juice of cell vacuole; ρ2 - interstitial fluid; ρ3-juice after grinding of tissue; ρ4-juice after plasmolysis ground tissue; ρ5-tissues of plant raw material, and the concentration ratio: kb - intercellular fluid in the tissue; kc - juice after grinding tissue; kd - juice after electroplasmolysis ground tissue. C - capacity of the cell membrane, mF and DU - controlled diode.

### III. CONCLUSION

1. Found that the resistivity of the plant tissue is one of the main physical characteristics of its state under electric processing.

2. Developed equivalent electrical equivalent circuit: cells; tissue ground tissue, chopped electroplasmolysed tissue
3. Obtained formulas for determining of the impedance: cells, tissue, ground tissue, ground electroplasmolysed tissue;
4. Derived formulas for determining the resistivity: cells; tissue ground tissue, chopped electroplasmolysed tissue.
5. Measured resistivity: sap vacuole cells, interstitial fluid, tissue plant material, tissue plant material after grinding, tissue plant material after grinding and electroplasmolysis;
6. Determined by the concentration ratio: interstitial fluid in tissue juice after grinding tissue juice after electroplasmolysis ground tissue.
7. Tracings were obtained cross-sectional areas of the protoplast and the cell wall in control and after plasmolysis, which showed that during electroplasmolysis undergoing profound changes in the cytoplasm of cells, violating the integrity of plasma membranes - plasma membrane and tonoplast.

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